

# HIV-Specific CD4<sup>+</sup> T Cells May Contribute to Viral Persistence in HIV Controllers

Peter W. Hunt,<sup>1,2</sup> Hiroyu Hatano,<sup>1,2</sup> Elizabeth Sinclair,<sup>1,2</sup> Tzong-Hae Lee,<sup>3</sup> Michael P. Busch,<sup>1,2,3</sup> Jeffrey N. Martin,<sup>1,2</sup> Joseph M. McCune,<sup>1,2</sup> and Steven G. Deeks<sup>1,2</sup>

<sup>1</sup>Departments of Medicine and <sup>2</sup>Laboratory Medicine, University of California; and <sup>3</sup>Blood Systems Research Institute, San Francisco, California

**Background.** Human immunodeficiency virus (HIV)--infected individuals maintaining plasma HIV RNA levels <75 copies/mL in the absence of therapy ("HIV controllers") often maintain high HIV-specific T cell responses, which likely contribute to the control of viral replication. Despite robust immune responses, these individuals never eradicate HIV infection. We hypothesized that HIV-specific CD4<sup>+</sup> T cells might serve as target cells for HIV, contributing to viral persistence in this setting.

**Methods.** We measured frequencies of activated (CD38<sup>+</sup> HLA-DR<sup>+</sup>) and HIV Gag-specific CD4<sup>+</sup> and CD8<sup>+</sup> T cells and plasma- and cell-associated levels of HIV RNA and DNA in a cohort of 38 HIV controllers.

**Results.** Although there was no evidence of a relationship between the extent of low-level viremia and the frequency of either activated or HIV-specific CD4<sup>+</sup> T cells, controllers with higher HIV-specific CD4<sup>+</sup> T cell frequencies had higher cell-associated HIV DNA levels ( $\rho = 0.53$ ;  $P = .019$ ). Higher activated CD4<sup>+</sup> T cell frequencies were also associated with higher levels of cell-associated DNA ( $P = .027$ ) and RNA ( $P = .0096$ ). However, there was no evidence of a relationship between cell-associated HIV RNA or DNA levels and HIV-specific CD8<sup>+</sup> T cell frequencies.

**Conclusions.** These data support a model in which strong HIV-specific CD4<sup>+</sup> T cell responses in HIV controllers, while contributing to a potent adaptive immune response, may also contribute to viral persistence, preventing the natural eradication of HIV infection.

Intense interest has been focused on "human immunodeficiency virus (HIV) controllers," a rare group of HIV-infected individuals who maintain clinically undetectable plasma HIV RNA levels (<75 copies/mL) in the absence of antiretroviral therapy. There is now broad consensus that highly functional HIV-specific T cells play an important role in the suppression of viral replication in most of these individuals [1–13]—observations that have supported the development of T cell immunity vaccines for HIV. Because some of these individuals have maintained both clinically undetectable viremia and normal CD4<sup>+</sup> T cell counts for >2 decades, many have also hypothesized that harnessing the

potent HIV-specific T cell responses observed in these individuals might allow for a "functional cure" of HIV infection. However, a recent T cell-mediated immunity vaccine for HIV failed to prevent HIV acquisition and actually increased the risk of infection in uncircumcised men [14]. There may also be negative inflammatory consequences to the immunologic control of HIV infection in HIV controllers, including systemic immune activation—occasionally contributing to significant decreases in the CD4<sup>+</sup> T cell count and to AIDS—and atherosclerosis [15, 16]. Finally, despite potent HIV-specific T cell responses and relatively small reservoirs of latently infected cells, no HIV controller has ever completely eradicated HIV [17–19].

It remains unclear why HIV controllers never succeed in completely eradicating HIV. One possibility is that the very immune response critical for the immunologic control of viral replication paradoxically promotes HIV persistence. For example, HIV-specific CD4<sup>+</sup> T cells are highly susceptible targets for direct HIV infection, because they preferentially activate and expand at sites of

Received 8 August 2010; accepted 3 December 2010.

Correspondence: Peter W. Hunt, MD, UCSF Positive Health Program, SFGH Bldg 80, Ward 84, 995 Potrero Ave, San Francisco, CA 94110 (phunt@php.ucsf.edu).

**Clinical Infectious Diseases** 2011;52(5):681–687

© The Author 2011. Published by Oxford University Press on behalf of the Infectious Diseases Society of America. All rights reserved. For Permissions, please e-mail: journals.permissions@oup.com.

1058-4838/2011/525-0001\$37.00

DOI: 10.1093/cid/ciq202

HIV replication [20]. We hypothesized that the high frequencies of activated and HIV-specific CD4<sup>+</sup> T cells observed in HIV controllers [3, 5, 6, 16] might actually serve as primary target cells, continuously replenishing the reservoir of latently infected cells and preventing complete eradication in these individuals. To begin to address this hypothesis, we assessed the relationship between activated and HIV-specific T cell responses and both plasma and cell-associated HIV RNA and DNA levels in a cohort of HIV controllers.

## MATERIALS AND METHODS

### Participants

HIV-infected adults were sampled from the Study of the Consequences of the Protease Inhibitor Era (SCOPE), a clinic-based cohort of >1000 HIV-infected individuals at the University of California, San Francisco. From this cohort, we sampled HIV controllers, defined as HIV-seropositive individuals maintaining clinically undetectable plasma HIV RNA levels (<75 copies/mL by bDNA or <50 copies/mL by polymerase chain reaction) for ≥5 years in the absence of therapy, with ≥2 plasma HIV RNA determinations in a given 1-year period. Isolated episodes of clinically detectable viremia up to 1000 copies/mL were allowed if they were followed by undetectable values. Plasma- and cell-associated levels of HIV RNA and DNA in these individuals were reported recently [19]. A subset of these individuals had cellular immunology studies performed and are the basis of the current report. All participants provided informed consent, and this research was approved by the institutional review board of the University of California, San Francisco.

### Laboratory Studies

**T Cell Activation** Freshly collected whole-blood samples were assessed for the frequency of activated (CD38<sup>+</sup>/HLA-DR<sup>+</sup>) CD4<sup>+</sup> and CD8<sup>+</sup> T cells using 4-color flow cytometry, setting quadrant gates on isotype controls to define positive and negative populations, as described elsewhere [16]. T cell activation was assessed as the percentage of CD4<sup>+</sup> and CD8<sup>+</sup> T cells coexpressing HLA-DR and CD38, because these measures have been associated with clinical progression in untreated individuals—even among HIV controllers—and poor CD4<sup>+</sup> T cell recovery in the presence of antiretroviral therapy [16, 21, 22].

**Cytokine Flow Cytometry** Fresh whole-blood samples were stimulated with overlapping peptide pools of HIV-1 p55 Gag or cytomegalovirus (CMV) pp65 protein (BD Biosciences) for 6 h, then fixed, permeabilized, and assessed for interferon (IFN)- $\gamma$  and interleukin (IL)-2 expression by CD4<sup>+</sup> and CD8<sup>+</sup> T cells, as previously described (gating strategy depicted in Figure 1 of reference [9]). Cytokine secretion in an unstimulated control

culture was subtracted to correct for nonspecific cytokine secretion. The frequencies of Gag-specific CD4<sup>+</sup> and CD8<sup>+</sup> T cells secreting both IFN- $\gamma$  and IL-2 were used as our primary measures of the Gag-specific T cell response, because these cells are more highly enriched in HIV controllers than those producing IFN- $\gamma$  alone and are believed to be more potent mediators of viral control [5, 9].

**Ultra-Sensitive Plasma HIV RNA Levels** Plasma HIV RNA levels were measured using an isothermal transcription mediated amplification (TMA) assay (Aptima; Gen-Probe) [23], with a 50% detection limit of 3.6–14 copies/mL when performed in singlicate [24]. The assay was performed in quadruplicate on 2 mL of plasma (obtained from the same time point as the T cell activation assays), improving the overall limit of detection to <3.5 copies/mL. The output is a ratio of signal to cutoff (S/Co; range, 0–30), with an S/Co ratio <1.0 considered undetectable [19].

**Cell-Associated HIV RNA and DNA Levels** The TMA assay was also used to measure cell-associated HIV RNA in cryopreserved peripheral blood mononuclear cells (PBMCs), using a modification of methods for PBMC extraction and TMA amplification of hepatitis C virus [25]. S/Co ratios were normalized to the input number of PBMCs. Total proviral HIV DNA was extracted from PBMCs using published methods and normalized to the input number of PBMCs [26]. This assay has an overall sensitivity of 1 copy/3  $\mu$ g of input DNA (~450,000 PBMCs) [27].

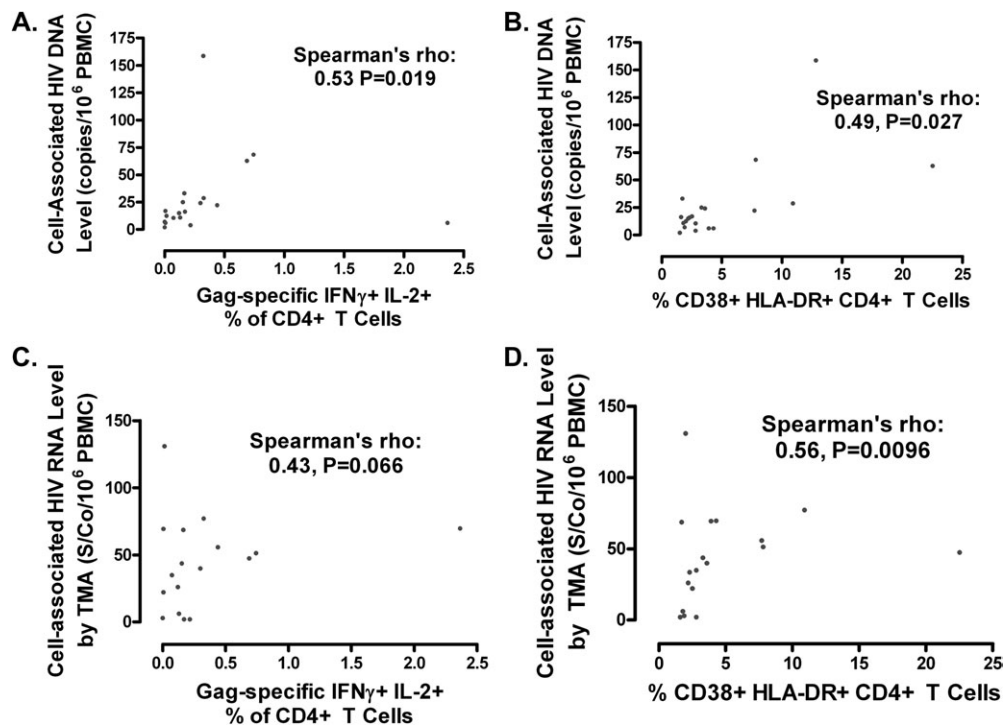
### Statistical Methods

Relationships between continuous variables were assessed with Spearman rank order correlation coefficients. Adjusted associations were assessed with linear regression, including potential confounders associated with the outcome of interest in unadjusted models ( $P > .10$ ) and calculating standard errors with heteroskedasticity-consistent covariance matrix estimators [28]. Models were considered valid only if inferences were robust to exclusion of influential outliers and leverage points.

## RESULTS

### Characteristics of Participants

A total of 38 HIV controllers with plasma HIV RNA levels <75 copies/mL in the absence of antiretroviral therapy and available cellular immunology data contributed to these analyses. Most were 40–50 years of age, and nearly one-half were women (Table 1). Most had received a diagnosis of HIV infection >1 decade previously, and most had normal CD4<sup>+</sup> T cell counts (median, 746 cells/mm<sup>3</sup>; interquartile range, 564–1037 cells/mm<sup>3</sup>). Only 3 HIV controllers (7%) had a CD4<sup>+</sup> T cell count <350 cells/mm<sup>3</sup>. Fourteen (37%) had at least 1 HLA B\*57 allele.



**Figure 1.** Strong relationship between cell-associated human immunodeficiency virus (HIV) burden and the frequency of activated and HIV-specific CD4<sup>+</sup> T cells in HIV controllers. The association between cell-associated HIV DNA and RNA levels and the frequencies of activated (CD38<sup>+</sup> HLA-DR<sup>+</sup>) and HIV Gag-specific CD4<sup>+</sup> T cells was assessed in peripheral blood mononuclear cells (PBMCs) from 19 HIV controllers. Higher cell-associated HIV DNA levels were strongly associated with higher frequencies of Gag-specific CD4<sup>+</sup> T cells (A) and activated CD4<sup>+</sup> T cells (B). Similarly, higher cell-associated HIV RNA levels were associated with higher frequencies of activated CD4<sup>+</sup> T cells (D) and tended to be associated with higher frequencies of Gag-specific CD4<sup>+</sup> T cells (C). IFN, interferon; IL, interleukin; S/Co, ratio of signal to cutoff; TMA, transcription mediated amplification.

### Weak and Inconsistent Relationships between the Extent of Low-Level Viremia and the Frequency of Activated and HIV-Specific T Cells

The extent of low-level viremia was assessed in all 38 HIV controllers using a high-sensitivity TMA assay. As previously reported, although all HIV controllers had undetectable plasma HIV RNA levels, as determined using clinically available assays, most HIV controllers had detectable HIV RNA in plasma by TMA (median S/Co ratio, 7.1; interquartile range, 1.8–18.8) [19]. Because the extent of low-level viremia might be a major determinant of the frequency of activated and HIV-specific T cells in these individuals, we assessed the relationship between these measurements. Although higher plasma HIV RNA levels

were modestly correlated with higher frequencies of activated (CD38<sup>+</sup> HLA-DR<sup>+</sup>) CD8<sup>+</sup> T cells ( $\rho = 0.32$ ;  $P = .048$ ), there was no evidence for a relationship between the extent of low-level viremia and the frequencies of activated CD4<sup>+</sup> T cells ( $\rho = 0.12$ ;  $P = .48$ ), HIV Gag-specific IFN $\gamma$ +IL-2+ CD4<sup>+</sup> T cells ( $\rho = -0.1$ ;  $P = .73$ ), or HIV Gag-specific IFN $\gamma$ +IL-2+ CD8<sup>+</sup> T cells ( $\rho = -0.04$ ;  $P = .81$ ).

### Strong Relationship between Activated and HIV-Specific CD4<sup>+</sup> T Frequencies and Cell-Associated HIV Burden

Although the extent of low-level viremia did not appear to be a major determinant of the frequency of activated and HIV-specific CD4<sup>+</sup> T cells, we hypothesized that the HIV-specific and activated

**Table 1. Characteristics of Human Immunodeficiency Virus (HIV) Controllers**

Characteristic	Data (n = 38)
Age, median years (IQR)	47 (44–50)
No. (%) of female subjects	16 (42)
CD4 cell count, median cells/mm <sup>3</sup> (IQR)	746 (564–1037)
Plasma HIV RNA level (branched DNA assay), median log <sub>10</sub> copies/mL	<75
Years of diagnosis of HIV infection	14 (8–18)
No. (%) of subjects with HLA B57 <sup>+</sup>	14 (37)

CD4<sup>+</sup> T cell expansion observed in most HIV controllers might increase the availability of target cells for HIV infection, contributing to viral persistence. To address this, we assessed the relationship between the activated and HIV-specific CD4<sup>+</sup> T cell frequencies and cell-associated reservoirs of HIV RNA and DNA in PBMCs in a subset of 18 HIV controllers. As expected, higher cell-associated HIV DNA levels were strongly associated with higher frequencies of HIV Gag-specific IFN- $\gamma$ +IL-2+ CD4<sup>+</sup> T cells ( $\rho = 0.53$ ;  $P = .019$ , Figure 1A) and activated CD4<sup>+</sup> T cells ( $\rho = 0.49$ ;  $P = .027$ ) (Figure 1B). Similarly, higher cell-associated HIV RNA levels were associated with higher frequencies of activated CD4<sup>+</sup> T cells ( $\rho = 0.56$ ;  $P = .0096$ ) (Figure 1D) and tended to be associated with higher frequencies of Gag-specific IFN- $\gamma$ +IL-2+ CD4<sup>+</sup> T cells ( $\rho = 0.43$ ;  $P = .066$ ) (Figure 1C). Similar trends were observed when the Gag-specific IFN- $\gamma$ + (with or without IL-2) CD4<sup>+</sup> T cell frequency was used to quantify HIV-specific CD4<sup>+</sup> T cells and when HLA-DR+ (with or without CD38) CD4<sup>+</sup> T cells was used to assess activated CD4<sup>+</sup> T cells. Confirming that the relationship between HIV-specific CD4<sup>+</sup> T cells and cellular HIV burden was antigen specific, there was no evidence for an association between cell-associated HIV RNA or DNA levels and CMV pp65-specific IFN- $\gamma$ +IL-2+ CD4<sup>+</sup> T cell frequencies ( $P > .27$  for both).

We also assessed for potential confounders of these relationships. Although there was no evidence of an association between age, sex, CD4 cell count, or HLA B\*57 status and cell-associated HIV DNA or RNA levels (all  $P > .10$ ), longer duration of HIV diagnosis (a variable underestimate of the duration of HIV infection) was associated with higher cell-associated HIV DNA levels ( $\rho = 0.47$ ;  $P = .048$ ). However, there was no evidence of a relationship between time since HIV diagnosis and activated or HIV-specific CD4<sup>+</sup> T cell frequencies ( $P > .59$  for both). Furthermore, after adjustment for time since HIV diagnosis, the activated and HIV-specific CD4<sup>+</sup> T cell frequencies continued to be strongly associated with cell-associated DNA levels ( $P < .001$  for both). It should be noted, however, that 2 influential outliers (one with the highest HIV DNA level and one with the highest HIV-specific CD4<sup>+</sup> T cell frequency in Figure 1A) needed to be excluded from the HIV-specific CD4<sup>+</sup> T cell model, because they acted as disproportionately influential leverage points that prevented modeling a linear relationship.

#### No Evidence of a Relationship between Activated and HIV-Specific CD8<sup>+</sup> T Cell Frequencies and Cell-Associated HIV Burden

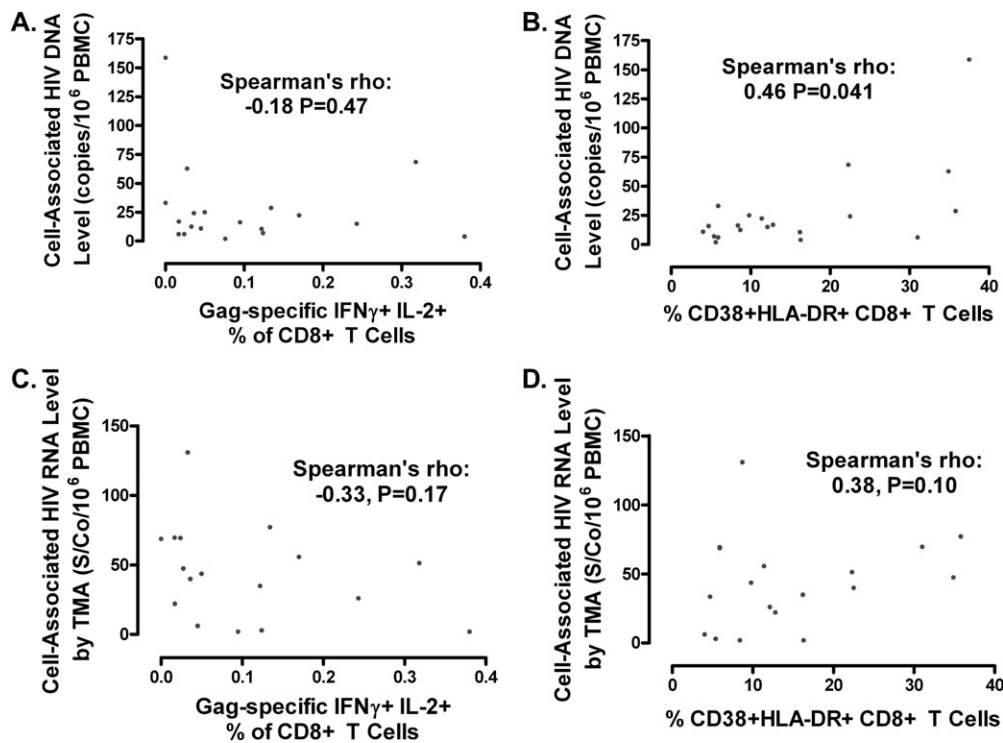
Because higher cellular burden of HIV could potentially be a cause rather than a consequence of activated and HIV-specific CD4<sup>+</sup> T cell expansion, we also assessed the relationship between cell-associated HIV burden and the frequency of activated and HIV-specific CD8<sup>+</sup> T cell responses. If a higher cell-associated burden of HIV was causing greater expansion of activated

and HIV-specific CD4<sup>+</sup> T cells, we would also expect to see a strong relationship between cellular HIV burden and activated and HIV-specific CD8<sup>+</sup> T cells. However, cell-associated HIV burden was less consistently associated with HIV-specific and activated CD8<sup>+</sup> T cells. Although cell-associated HIV DNA levels were associated with activated CD8<sup>+</sup> T cells ( $\rho = 0.46$ ;  $P = .041$ ) (Figure 2B), there was no evidence of an association with HIV-specific IFN- $\gamma$ +IL-2+ CD8<sup>+</sup> T cells ( $\rho = -0.18$ ,  $P = .47$ , Figure 2A). There was also no evidence for an association between cell-associated HIV RNA levels and frequencies of either HIV-specific IFN- $\gamma$ +IL-2+ CD8<sup>+</sup> T cells ( $\rho = -0.33$ ;  $P = .17$ ) (Figure 2C) or activated CD8<sup>+</sup> T cells ( $\rho = 0.38$ ;  $P = .10$ ) (Figure 2D). There was also no evidence of an association between the frequency of Gag-specific IFN- $\gamma$ + (with or without IL-2) CD8<sup>+</sup> T cells and either cell-associated HIV RNA or DNA levels ( $P > .45$  for both).

## DISCUSSION

Although most HIV controllers maintain strong HIV-specific T cell responses and extremely low levels of virus replication in the absence of therapy, no controller has ever eradicated HIV. We hypothesized that HIV-specific CD4<sup>+</sup> T cells, although potentially supporting an effective HIV-specific immune response, might also serve as major target cells for HIV, continually replenishing the reservoir of latently infected cells. Supporting this hypothesis, we found that HIV controllers with the highest frequencies of activated and HIV-specific CD4<sup>+</sup> T cells had the highest cell-associated burdens of HIV. Although higher viral burden could also be a cause of greater activated and HIV-specific CD4<sup>+</sup> T cell expansion, we observed no evidence of a relationship between the degree of low-level viremia and the frequency of these cells. Furthermore, we found no evidence of a relationship between the cell-associated viral burden and HIV-specific CD8<sup>+</sup> T cell frequencies. Collectively, these data support a model in which HIV-specific CD4<sup>+</sup> T cells may replenish the latent reservoir and prevent the natural eradication of HIV infection in HIV controllers.

Our results are consistent with a recent report demonstrating that CD4<sup>+</sup> T cells from HIV controllers are just as easily infected with HIV after ex vivo stimulation as cells from HIV-uninfected individuals [29]. Although viruses from HIV controllers may be somewhat less fit than viruses from viremic individuals [30, 31], replication-competent virus can be isolated from many controllers and some have been infected with highly pathogenic viruses from individuals with AIDS [32–34]. Despite infection with replication-competent viruses and having susceptible CD4<sup>+</sup> T cells, HIV controllers maintain much lower cell-associated DNA levels than do HIV-infected individuals maintaining treatment-mediated viral suppression [17–19]. These exceptionally small cellular reservoirs in HIV controllers have been



**Figure 2.** Weak and inconsistent relationships between cell-associated human immunodeficiency virus (HIV) burden and the frequency of activated and HIV-specific CD8<sup>+</sup> T cells in HIV controllers. The association between cell-associated HIV DNA and RNA levels and the frequencies of activated and HIV Gag-specific CD8<sup>+</sup> T cells was assessed in peripheral blood mononuclear cells (PBMCs) from 19 HIV controllers. Although higher frequencies of activated CD8<sup>+</sup> T cells were associated with higher cell-associated HIV DNA levels (B), there was no evidence of a relationship between the frequency of HIV Gag-specific CD8<sup>+</sup> T cells and cell-associated HIV DNA levels (A). There was also no evidence for a relationship between either HIV Gag-specific (C) or activated (D) CD8<sup>+</sup> T cell frequencies and cell-associated HIV RNA levels. IFN, interferon; IL, interleukin; S/Co, ratio of signal to cutoff; TMA, transcription mediated amplification.

largely attributed to a highly potent cytotoxic HIV-specific CD8<sup>+</sup> T cell response, observable in the majority of these individuals [1–13]. Enrichment for certain HLA and KIR allotypes in HIV controllers has also implicated a potential role of natural killer cells in clearing virally infected cells [35]. Despite these potent cytotoxic antiviral responses, HIV controllers never completely eradicate HIV. Our results are consistent with the hypothesis that the very HIV-specific CD4<sup>+</sup> T cell response coordinating an effective antiviral immune response may be contributing to viral persistence in this setting.

It has long been known that HIV-specific CD4<sup>+</sup> T cells are more likely to be infected with HIV than non-HIV-specific CD4<sup>+</sup> T cells [20]. It is worth carefully considering this point in the context of HIV controllers because—as a group—they harbor extremely high frequencies of HIV-specific CD4<sup>+</sup> T cells while maintaining extremely low proviral HIV DNA levels [3, 5, 6, 17–19]. Taken together, these observations might suggest that the latent reservoir in HIV controllers is more highly concentrated in the HIV-specific CD4<sup>+</sup> T cell population than in most other HIV-infected individuals. This might be the case if HIV controllers were primarily containing viral replication in

lymphoid tissues with high local concentrations of expanded HIV-specific T cells. If this hypothesis were true, it might have important implications for eradication strategies. For example, if reinfection of HIV-specific CD4<sup>+</sup> T cells could be prevented in HIV controllers with combination antiretroviral therapy—preferably composed of agents targeting preintegration steps—repeated therapeutic vaccination with HIV antigens could potentially stimulate virus production by latently infected HIV-specific CD4<sup>+</sup> T cells, allowing their clearance by the potent antiviral responses maintained by most of these individuals, potentially reducing the latent reservoir to low-enough levels that eradication might be possible. Although such a strategy would not have immediate relevance to the majority of HIV-infected individuals who fail to spontaneously control HIV replication, it might provide proof of principle that harnessing host anti-viral immune responses could play a role in HIV eradication.

The recognition that HIV-specific CD4<sup>+</sup> T cells are likely to be major target cells for HIV even in controllers may also have relevance for T cell immunity vaccine strategies for HIV. Vaccines that elicit strong proliferative HIV-specific CD4<sup>+</sup> T cell



responses might actually increase the risk of HIV acquisition, as appeared to be the case in the recent STEP trial [14]. Similarly, a therapeutic vaccine inducing a strong CD4<sup>+</sup> T cell response actually shortened the time to viral rebound among HIV-infected individuals undergoing treatment interruption [36], though this latter observation has not been observed with all therapeutic HIV vaccines [37]. Indeed, low systemic immune activation levels have been consistently associated with a decreased risk of HIV acquisition in highly exposed HIV-uninfected individuals [38–42], and blocking the recruitment of activated CD4<sup>+</sup> T cell targets to mucosal surfaces with a topical microbicide appeared to protect macaques from systemic infection by the simian immunodeficiency virus following vaginal challenge [43].

Our study has several limitations that deserve comment. First, our study is correlative, and we do not provide definitive proof that activated and HIV-specific CD4<sup>+</sup> T cells are major contributors to viral persistence in HIV controllers. Indeed, higher cell-associated viral burden could be a cause rather than a consequence of the expansion of activated and HIV-specific CD4<sup>+</sup> T cells observed. However, we found no evidence of an association between the extent of low-level viremia and the expansion of these cells. Furthermore, if higher viral burden was a major determinant of activated and HIV-specific T cell expansion in HIV controllers, we might have also expected to see strong and consistent associations between cell-associated viral burden and the frequency of HIV-specific CD8<sup>+</sup> T cells. However, we saw, at best, inconsistent evidence of these relationships. The lack of consistent relationships between cell-associated viral burden and activated and HIV-specific CD8<sup>+</sup> T cell responses might be explained by the countervailing antiviral activity of these cells. However, the contrasting relationships between cell-associated viral burden and HIV-specific CD4<sup>+</sup> and CD8<sup>+</sup> T cells further highlights the relative failure of HIV-specific CD4<sup>+</sup> T cells to prevent viral persistence. Ultimately, isolating CD4<sup>+</sup> T cells from blood and tissues and measuring HIV DNA levels separately within HIV-specific and non-HIV-specific fractions would be the most definitive way to assess whether the cellular viral burden in HIV controllers is preferentially maintained in the HIV-specific population [20], but this is beyond the scope of the current study. Finally, not every HIV controller with high HIV-specific CD4<sup>+</sup> T cell frequencies in our sample had high HIV DNA levels. In fact, the controller with the highest HIV-specific CD4<sup>+</sup> T cell frequency had a very low HIV DNA level. Additional testing is necessary to address whether this individual has favorable CCR5 genetics or other important cellular restriction factors.

In summary, although most HIV controllers maintain high frequencies of potent HIV-specific T cells, none has ever successfully eradicated HIV. The strong relationship between the frequency of activated and HIV-specific CD4<sup>+</sup> T cells and cell-

associated viral burden in controllers may suggest that these cells are continually replenishing the latent reservoir, preventing the natural eradication of HIV in this setting. These observations have important implications both for the design of effective HIV vaccines and microbicides and for novel HIV eradication strategies.

## Acknowledgments

**Financial support.** This work was supported in part by the UCSF/Gladstone Center for AIDS Research (P30 AI27763, P30 MH59037), NIAID (AI065244, AI44595, and AI-76174), the Center for AIDS Prevention Studies (P30 MH62246), the UCSF Clinical and Translational Science Institute (UL1 RR024131-01), the CFAR Network of Integrated Clinical Sciences (5R24AI067039), the Ragon Institute (of MGH, MIT, and Harvard), and American Foundation for AIDS Research (106710-40-RGRL). J.M.M. is a recipient of a grant (DPI OD00329) from the National Institutes of Health's (NIH) Director's Pioneer Award Program, part of the NIH Roadmap for Medical Research.

**Potential conflicts of interest.** M.P.B. has consulted for Gen-Probe and Creative Testing Solutions; received grant support from the Novartis/Chiron and Caridian BCT; has received Payment for lectures or has served on the speakers' bureau for Ortho Diagnostics, Roche Molecular Diagnostics, Novartis/Chiron, Abbott Laboratories, and BioRad; and has had travel/meeting expenses paid by the AABB, ISBT, IPFA, WHO, CDC, FDA, NIH.

## References

- Betts MR, Nason MC, West SM, et al. HIV nonprogressors preferentially maintain highly functional HIV-specific CD8<sup>+</sup> T cells. *Blood* 2006; 107:4781–9.
- Migueles SA, Osborne CM, Royce C, et al. Lytic granule loading of CD8(+) T cells is required for HIV-infected cell elimination associated with immune control. *Immunity* 2008; 29:1009–21.
- Potter SJ, Lacabaratz C, Lambotte O, et al. Preserved central memory and activated effector memory CD4<sup>+</sup> T-cell subsets in human immunodeficiency virus controllers: an ANRS EP36 study. *J Virol* 2007; 81:13904–15.
- Bailey JR, Brennan TP, O'Connell KA, Siliciano RF, Blankson JN. Evidence of CD8<sup>+</sup> T-cell-mediated selective pressure on human immunodeficiency virus type 1 nef in HLA-B\*57<sup>+</sup> elite suppressors. *J Virol* 2009; 83:88–97.
- Emu B, Sinclair E, Favre D, et al. Phenotypic, functional, and kinetic parameters associated with apparent T-cell control of human immunodeficiency virus replication in individuals with and without antiretroviral treatment. *J Virol* 2005; 79:14169–78.
- Harari A, Petitpierre S, Vallelian F, Pantaleo G. Skewed representation of functionally distinct populations of virus-specific CD4 T cells in HIV-1-infected subjects with progressive disease: changes after antiretroviral therapy. *Blood* 2004; 103:966–72.
- Saez-Cirion A, Lacabaratz C, Lambotte O, et al. HIV controllers exhibit potent CD8 T cell capacity to suppress HIV infection ex vivo and peculiar cytotoxic T lymphocyte activation phenotype. *Proc Natl Acad Sci U S A* 2007; 104:6776–81.
- Pereyra F, Addo MM, Kaufmann DE, et al. Genetic and immunologic heterogeneity among persons who control HIV infection in the absence of therapy. *J Infect Dis* 2008; 197:563–71.
- Emu B, Sinclair E, Hatano H, et al. HLA class I-restricted T-cell responses may contribute to the control of human immunodeficiency virus infection, but such responses are not always necessary for long-term virus control. *J Virol* 2008; 82:5398–407.
- van Manen D, Kootstra NA, Boeser-Nunnink B, Handulle MA, van't Wout AB, Schuitemaker H. Association of HLA-C and HCP5 gene regions with the clinical course of HIV-1 infection. *AIDS* 2009; 23:19–28.

11. Fellay J, Shianna KV, Ge D, et al. A whole-genome association study of major determinants for host control of HIV-1. *Science* **2007**; 317: 944–7.
12. Catano G, Kulkarni H, He W, et al. HIV-1 disease-influencing effects associated with ZNRD1, HCP5 and HLA-C alleles are attributable mainly to either HLA-A10 or HLA-B\*57 alleles. *LoS One* **2008**; 3:e3636.
13. Limou S, Le Clerc S, Coulonges C, et al. Genomewide association study of an AIDS-nonprogression cohort emphasizes the role played by HLA genes (ANRS Genomewide Association Study 02). *J Infect Dis* **2009**; 199:419–26.
14. Buchbinder SP, Mehrotra DV, Duerr A, et al. Efficacy assessment of a cell-mediated immunity HIV-1 vaccine (the Step Study): a double-blind, randomised, placebo-controlled, test-of-concept trial. *Lancet* **2008**; 372:1881–93.
15. Hsue PY, Hunt PW, Schnell A, et al. Role of viral replication, antiretroviral therapy, and immunodeficiency in HIV-associated atherosclerosis. *AIDS* **2009**; 23:1059–67.
16. Hunt PW, Brechley J, Sinclair E, et al. Relationship between T cell activation and CD4(+) T cell count in HIV-seropositive individuals with undetectable plasma HIV RNA levels in the absence of therapy. *J Infect Dis* **2008**; 197:126–33.
17. Garbuglia AR, Salvi R, Di Caro A, et al. Peripheral lymphocytes of clinically non-progressor patients harbor inactive and uninducible HIV proviruses. *J Med Virol* **1995**; 46:116–21.
18. Lambotte O, Boufassa F, Madec Y, et al. HIV controllers: a homogeneous group of HIV-1-infected patients with spontaneous control of viral replication. *Clin Infect Dis* **2005**; 41:1053–6.
19. Hatano H, Delwart EL, Norris PJ, et al. Evidence for persistent low-level viremia in individuals who control human immunodeficiency virus in the absence of antiretroviral therapy. *J Virol* **2009**; 83:329–35.
20. Douek DC, Brechley JM, Betts MR, et al. HIV preferentially infects HIV-specific CD4<sup>+</sup> T cells. *Nature* **2002**; 417:95–8.
21. Carbone J, Gil J, Benito JM, et al. Increased levels of activated subsets of CD4 T cells add to the prognostic value of low CD4 T cell counts in a cohort of HIV-infected drug users. *AIDS* **2000**; 14:2823–9.
22. Hunt PW, Martin JN, Sinclair E, et al. T cell activation is associated with lower CD4<sup>+</sup> T cell gains in human immunodeficiency virus-infected patients with sustained viral suppression during antiretroviral therapy. *J Infect Dis* **2003**; 187:1534–43.
23. Stramer SL, Glynn SA, Kleinman SH, et al. Detection of HIV-1 and HCV infections among antibody-negative blood donors by nucleic acid-amplification testing. *N Engl J Med* **2004**; 351:760–8.
24. Busch MP, Glynn SA, Wright DJ, et al. Relative sensitivities of licensed nucleic acid amplification tests for detection of viremia in early human immunodeficiency virus and hepatitis C virus infection. *Transfusion* **2005**; 45:1853–63.
25. Bernardin F, Tobler L, Walsh I, Williams JD, Busch M, Delwart E. Clearance of hepatitis C virus RNA from the peripheral blood mononuclear cells of blood donors who spontaneously or therapeutically control their plasma viremia. *Hepatology* **2008**; 47:1446–52.
26. Lee TH, el-Amad Z, Reis M, et al. Absence of HIV-1 DNA in high-risk seronegative individuals using high-input polymerase chain reaction. *AIDS* **1991**; 5:1201–7.
27. Lee TH, Paglieroni T, Utter GH, et al. High-level long-term white blood cell microchimerism after transfusion of leukoreduced blood components to patients resuscitated after severe traumatic injury. *Transfusion* **2005**; 45:1280–90.
28. Davidson R, MacKinnon J. *Estimation and inference in econometrics*. New York: Oxford University Press, 1993.
29. Julg B, Pereyra F, Buzon MJ, et al. Infrequent recovery of HIV from but robust exogenous infection of activated CD4(+) T cells in HIV elite controllers. *Clin Infect Dis* Jul **2010**; 51:233–8.
30. Lassen KG, Lobritz MA, Bailey JR, et al. Elite suppressor-derived HIV-1 envelope glycoproteins exhibit reduced entry efficiency and kinetics. *PLoS Pathog* **2009**; 5:e1000377.
31. Miura T, Brockman MA, Schneidewind A, et al. HLA-B57/B\*5801 HIV-1 elite controllers select for rare gag variants associated with reduced viral replication capacity and strong Ctl recognition. *J Virol* **2009**; 83:2743–55.
32. Lamine A, Caumont-Sarcos A, Chaix ML, et al. Replication-competent HIV strains infect HIV controllers despite undetectable viremia (ANRS EP36 study). *AIDS* **2007**; 21:1043–5.
33. Bailey JR, O'Connell K, Yang HC, et al. Transmission of human immunodeficiency virus type 1 from a patient who developed AIDS to an elite suppressor. *J Virol* **2008**; 82:7395–410.
34. Blankson JN, Bailey JR, Thayil S, et al. Isolation and characterization of replication-competent human immunodeficiency virus type 1 from a subset of elite suppressors. *J Virol* **2007**; 81:2508–18.
35. Martin MP, Qi Y, Gao X, et al. Innate partnership of HLA-B and KIR3DL1 subtypes against HIV-1. *Nat Genet* **2007**; 39:733–40.
36. Autran B, Murphy RL, Costagliola D, et al. Greater viral rebound and reduced time to resume antiretroviral therapy after therapeutic immunization with the ALVAC-HIV vaccine (vCP1452). *AIDS* **2008**; 22:1313–22.
37. Schooley RT, Spritzler J, Wang H, et al. AIDS clinical trials group 5197: a placebo-controlled trial of immunization of HIV-1-infected persons with a replication-deficient adenovirus type 5 vaccine expressing the HIV-1 core protein. *J Infect Dis* Sep **2002**; 705–16.
38. Koning FA, Otto SA, Hazenberg MD, et al. Low-level CD4<sup>+</sup> T cell activation is associated with low susceptibility to HIV-1 infection. *J Immunol* **2005**; 175:6117–22.
39. Begaud E, Chartier L, Marechal V, et al. Reduced CD4 T cell activation and in vitro susceptibility to HIV-1 infection in exposed uninfected Central Africans. *Retrovirology* **2006**; 3:35.
40. Jennes W, Evertse D, Borget MY, et al. Suppressed cellular alloimmune responses in HIV-exposed seronegative female sex workers. *Clin Exp Immunol* **2006**; 143:435–44.
41. Salkowitz JR, Purvis SF, Meyerson H, et al. Characterization of high-risk HIV-1 seronegative hemophiliacs. *Clin Immunol* **2001**; 98:200–11.
42. Card CM, McLaren PJ, Wachihhi C, Kimani J, Plummer FA, Fowke KR. Decreased immune activation in resistance to HIV-1 infection is associated with an elevated frequency of CD4(+)CD25(+)FOXP3(+) regulatory T cells. *J Infect Dis* **2009**; 199:1318–22.
43. Li Q, Estes JD, Schlievert PM, et al. Glycerol monolaurate prevents mucosal SIV transmission. *Nature* **2009**; 458:1034–8.